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Biosensors for Selective and Label-free Detection of *Xylella fastidiosa*

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The *Xylella fastidiosa* bacterium is one of the most hostile pathogenic microorganisms causing multiple plant diseases, with huge economic impact on both agriculture and environment. [1] By revealing the presence of the bacterium before symptom arrival, preventive action plans could be applied to confine the contagion. In this scenario, the development of a surveillance device, conveying high sensitivity along with early detection of *X. fastidiosa* outbreaks, would be of paramount importance. Fundamental to the development of such a detection system is the availability of high binding affinity antibodies for *X. fastidiosa* (anti-XF), which could be integrated in the sensor transducers. Also, a label-free detection system would be desirable, which shorts the analysis process to achieve optimal time-to-results for pathogen identification.[2]

In this study Surface Plasmon Resonance (SPR) has been used to assess an optimized biofunctionalization protocol of gold surfaces with anti-XF, validating their capturing efficacy against *X. fastidiosa*. [3] Among other bio-sensing techniques, SPR holds the advantage of being a label-free detection system, providing real-time monitoring of bio-affinity reactions.[4] So far, a variety of plant pathogen biomarkers were studied by means of SPR, but none of them involves *X. fastidiosa*. [5]

The bio-functionalization of gold transducing interfaces was performed with the polyclonal antibodies for *X. fastidiosa*, covalently bounded to a self-assembled monolayer of alkylthiols. This configuration guaranteed the assay selectivity, which is assessed by means of a control experiment with the non-binding *Burkholderia phytofirmans* bacterium. The SPR sensogram of the binding *X. fastidiosa* is depicted in Figure 1A, and the comparison with the control experiment response is reported in Figure 1B.

Remarkably, a limit of detection as low as 10^5 CFU/mL was achieved by transducing the direct interaction between the *X. fastidiosa* bacterium and its affinity antibody, which is comparable to the label-needing ELISA gold standard. Moreover, the binding-affinity between polyclonal antibodies and the *X. fastidiosa* bacteria has been also evaluated, obtaining an equilibrium affinity constant of $3.5 \cdot 10^7$ M⁻¹, comparable with those given in the literature for bacteria detection against affinity antibodies.[6] The study is therefore a preliminary development of a reliable cost-effective process to successfully bio-functionalize a gold surface, eventually suitable as gate electrode in wide-field bioelectronic sensors, for ultra-sensitive detection of *X. fastidiosa*. [7]

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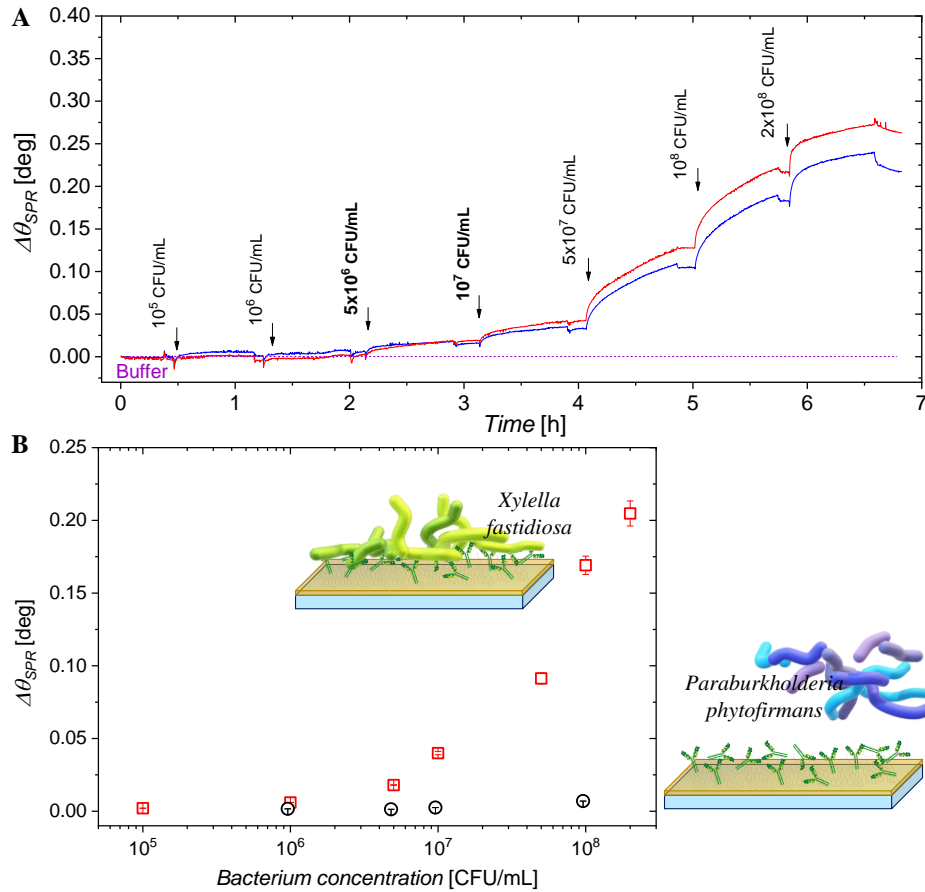


Fig.1 A) Exposure of the anti-XF functionalized surface to *X. fastidiosa* at increasing concentrations; black arrows correspond to sample injections and purple dotted line refers to the buffer level. Red and blue lines refer to two sampling points measured simultaneously. B) Comparison of *X. fastidiosa* (red square) and *Burkholderia phytofirmans* (black circle) SPR responses against the anti-XF functionalized surface. The average signal and standard deviation for four replicates analysis are reported.[3]

References

[1] C. Cariddi, M. Saponari, D. Boscia, A. De Stradis, G. Loconsole, F. Nigro, F. Porcelli, O. Potere, G. P. Martelli, Isolation of a *Xylella fastidiosa* strain infecting olive and oleander in Apulia, Italy. *J. Plant Pathol.* 96 (2014) 425.

[2] M. Khater, A.de la Escosura-Muñiz, A.Merkoçi, Biosensors for plant pathogen detection. *Biosensors and Bioelectronics* 93 (2017) 72–86.

[3] L. Sarcina, E. Macchia, G. Loconsole, G. D’Attoma, P. Saldarelli, V. Elicio, G. Palazzo and L. Torsi, Surface Plasmon Resonance assay for label-free and selective detection of *Xylella fastidiosa*. *Adv. NanoBiomed Res.* (2021) 2100043.

[4] V. Nanduri, A. K. Bhunia, S. I. Tu, G. C. Paoli, J. D. Brewster, SPR biosensor for the detection of *L. monocytogenes* using phage-displayed antibody. *Biosens. Bioelectron.* 23 (2007) 248.

[5] A. D. Taylor, J. Ladd, Q. Yu, S. Chen, J. Homola, S. Jiang, Quantitative and simultaneous detection of four foodborne bacterial pathogens with a multi-channel SPR sensor. *Biosens. Bioelectron.* 22 (2006) 752.

[6] E. Rostova, C. Ben Adiba, G. Dietler, S. K. Sekatskii, Kinetics of antibody binding to membranes of living bacteria measured by a photonic crystal-based biosensor. *Biosensors* 6 (2016) 52.

[7] E. Macchia, K. Manoli, B. Holzer, C. Di Franco, M. Ghittorelli, F. Torricelli, D. Alberga, G. F. Mangiatordi, G. Palazzo, G. Scamarcio, L. Torsi, Single-molecule detection with a millimetre-sized transistor. *Nat. Commun.* 9 (2018) 3223.